

A STUDY OF THE PHYTOCHEMICAL, ANTI-OXIDANT AND ANTI-INFLAMMATORY PROPERTIES OF CLOVES (*Syzygium Aromaticum*).

by

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Abstract

Syzygium aromaticum, commonly called clove, is a culinary spice with medical uses with medical uses and its main constituents possess antimicrobial, antioxidant, anti-inflammatory, analgesic, anticancer, and anesthetic effects. In this study, the antioxidant activity, anti-inflammatory activity and phytochemical constituent of *Syzygium aromaticum*'s flower bud were analyzed using three different extracts solvents (carbon tetrachloride, ethyl acetate and aqueous extract). The analyses were carried out using standard analytical methods. Phytochemical analysis was carried out using GC-FID while other parameter was assayed using spectrophotometer. The results showed that carbon tetrachloride extract possessed the highest phytochemical profile (22.311273ppm) while aqueous extract has the lowest phytochemical profile (9.8821797ppm). All the extract both in ABTS and H_2O_2 scavenging activity are significantly low ($p < 0.05$) when compared to their standards and as a result may not be a good anti-oxidant agents. Aqueous extract showed a strong lipid peroxidation scavenging activity with an average value of 0.2987 $\mu\text{mol/ml}$ while carbon tetrachloride has the least value of 0.1328 $\mu\text{mol/ml}$ when compared with each other. The highest DPPH capacity was shown by Carbon tetrachloride extract with the average value of 89.7966% and the least is 86.332% ethyl acetate extract when compared with each other. In anti-inflammatory activity, the result shows that aqueous and carbon tetrachloride extract are not significant ($p > 0.05$) mean they are good albumin anti-inflammatory agents while ethyl acetate is significant low ($p < 0.05$) mean not a good anti-inflammatory agent when compared with aspirin (standard). In Heat induced Hemolysis, ethyl acetate extract is the only good anti-inflammatory agent because it is the only not significant extract ($p > 0.05$) when compared with the standard while the other are significant ($p < 0.05$). Carbon tetrachloride extract showed a strong Anti-proteinase activity because it is the only not significant extracts ($p > 0.05$) when compared with standard in this assay. Therefore, the study shows that all the extract of clove *syzyium aromaticum* may not possess the better anti-oxidant properties while in anti- inflammatory property carbon tetrachloride extract possess a better anti-inflammatory property.

Keywords: Phytochemical, Anti-oxidant, Anti-inflammatory, Properties & Clove

Introduction

Over the last decades, aromatic plants have been used traditionally as food preservative and for many therapeutic purposes (Shahid *et al*, 2019). Nowadays, the therapeutic properties of aromatic plants are used in the modern medicine; the volatile compound of those aromatic plant are the common

ingredients of pharmaceutical (Dorsaf *et al*, 2021). So, this plant is important to human everyday life (Yadav *et al*, 2020). There are different kinds of aromatic plants such as Cloves (*Syzygium aromaticum*), Peppermint (*Mentha* spp), Lavender (*Lavandula* spp), Lemon balm (*Melissa officinalis*) and Rosemary (*Salvia rosmarinus*). Antimicrobial and antioxidant properties are essential for food preservation and medicine which are found in Cloves (*Syzygium aromaticum*). Cloves (*Syzygium aromaticum*) is a dried flower bud which belongs to Myrtaceae family and the color is small brown (Batiha *et al*, 2020). It is evergreen tropical Myrtaceae family tree native to the island of Maluka in east Indonesia and it's widely distributed in tropical and subtropical areas of Asia, Africa, Madagascar and throughout Pacific and Oceanic region (Batiha *et al*, 2020). It was used as food preservatives, flavoring agents and nutritional additives, medicinal coloring agents (Cortes-Rojas *et al*, 2014).

Phytochemicals are biologically active, naturally occurring chemical compounds found in plants which include saponins, alkaloids, tannins, flavonoid and phenolic compounds (Bacanli *et al*, 2019). The phytochemical analysis presence on the plant's extract reported the presence of eugenol, acetyl eugenol, Alpha and Beta-caryophyllene, vanillin, tannins in cloves. (Yusuf *et al*, 2021). Analysis of phytochemicals by GC-FID (Gas Chromatography-Flame Ionization Detector) is one of the modern techniques used to identify and isolate Phyto-constituents (Nwiloh *et al*, 2016). Cloves shows some pharmacological activities such as anti-oxidant, anti-cancer, anti-inflammatory, anti-pyretic, anti-viral, anti-diabetic, anesthetic, analgesic, anti-carcinogenic, antibacterial, antifungal, antibiotic (Yunusa *et al*, 2018).

Inflammation is a defense response of our body to hazardous stimuli such as allergens and/or injury to the tissues; on the other hand, uncontrolled inflammatory response is the main cause of a vast continuum of disorders

including allergies, cardiovascular dysfunctions, metabolic syndrome, cancer, and autoimmune disease imposing a huge economic burden on individuals and consequently on the society. Inflammation is a comprehensive array of physiological response to a foreign organism including human pathogens, dust particles and viruses, (Arulselvan *et al*, 2016). Inflammation can be created by several different causes including a blood clot, an immune system disorder, a cancer, an infection, a chemical exposure, a physical injury or a neurological condition, such as Alzheimer's or depression (Roe, 2021). It can also arise as a result of production of free radicals from various sources due to an imbalance of natural antioxidants.

Antioxidants are molecules that inhibit or quench free radical reactions and delay or inhibit cellular damage (Young *et al*, 2001). Antioxidants compounds act as "free radical scavenger" by preventing and repairing damages caused reactive oxygen species (ROS) and therefore enhance the immune defense and lower the risk of cancer and degenerative disease (Valko *et al*, 2006). Free radical and ROS are generated by living cells during respiration and other cellular activities and endogenous antioxidants scavenge these free radicals/ROS to prevent them from attacking biomolecules and causing damaging effects in the cell. Oxidative damage to biological systems exceeds the scavenging/quenching capacity of the cell's endogenous antioxidants. These may result in DNA damage or degradation of protein and induction of lipid peroxidation. (Kola, 2019).

This study evaluates the phytochemical, anti-inflammatory and anti-oxidants properties of aqueous, ethyl acetate and carbon tetrachloride extracts that were extracted from Cloves (*Syzygium aromaticum*).

Aim of Study

The aim of this study is to investigate the phytochemical properties, anti-inflammatory properties and the anti-oxidants properties of different solvent extracts that were extracted from *Syzygium aromaticum* (cloves).

Objective of Study

1. To determine the phytochemical properties of Syzygium aromaticum extracts
2. To determine the anti-inflammatory and identified by a taxonomist in the properties of Syzygium aromaticum; the department of Botany, Nnamdi Azikiwe University, Awka and it was bought in its albumin denaturation and the anti-proteinase activity on Syzygium aromaticum (clove) in each of the extracts.
3. To determine the anti-oxidant properties in Syzygium aromaticum; the ABTs scavenging effects, the hydrogen peroxide scavenging effects, assay of lipid peroxidation and DPPH scavenging effect in each of the extracts of Syzygium aromaticum (cloves).
4. To examine which among the three extracts has high phytochemical, anti-oxidant and anti-inflammatory properties
5. To examine if there is any significant difference in the three extracts when compared to standard.

Materials

1. Spectrophotometer
2. Weighing balance
3. Centrifuge
4. Hot air oven
5. Beakers
6. Aluminum foils
7. Measuring cylinder
8. Heating mantle
9. Pasteur pipette
10. Reagent bottles
11. Test tubes and rack
12. Masking tape
13. Funnel
14. Cuvette

Reagents

1. ABTS Solution (7mM with 2.45mM ammonium persulfate)
2. Phosphate buffer (0.1M, pH 7.4)
3. H_2O_2 (40mM) in phosphate buffer
4. TCA (10%)
5. TBA (0.1M)
6. Phosphate buffer (0.12M, pH 7.2)
7. DPPH – 2,2-diphenyl-2-picrylhydrazyl hydrate (0.3mM in methanol)
8. Methanol

Method of Sample Collection

The sample used for this project were bought at Eke-Awka market Awka, Anambra state and identified by a taxonomist in the department of Botany, Nnamdi Azikiwe University, Awka and it was bought in its dried form after which it was grinded into powder ready for work

Method of Sample Preparation

600g of sample were weighed and macerated in 1500ml of methanol. The sample mixture was extracted for 72hrs. After 72hrs, it was filtered using muslin cloth. The extract was filtered and the filtrate was finally dried at low room temperature (60°C) under pressure in a rotary vacuum evaporator (Thermotech, buchi type model th-012). The extracts were concentrated, percentage yield calculated and then subjected to column chromatography and the product of column subjected to phytochemical screening using GC-FID, anti-oxidants and anti-inflammatory analysis. The dried extract was properly stored in the desiccators for further experiment and analysis.

Method of Analysis

Column chromatography

1. 50g of Silica gel G was heated in an oven at 130°C for 4hrs and was transferred to a 250ml size beaker and placed in a desiccator.
2. 10g of silica gel was package into the conventional size chromatography column and a glass wool was plugged in the bottom. The column has a clamp to stop the flow of the solvent. And 10 ml of ethanol was mixed with 10 g silica to obtain slurry. The column was filled with the slurry until alumina settles down to 4-5 cm height.
3. Some ethanol was kept above the silica's top and the silica was not allowed to be dry. Ethanol was about 1 mm at the top of the column and a stopper at the bottom of the column with a hose clamp.
4. Crude extract was eluted with a step gradient of n-hexane, ethyl acetate and

methanol. N-hexane was first added to the crude extract and was properly mixed and then transferred to the column; The process was repeated until all the component of the extract soluble in n-hexane were extracted.

5. The process was repeated with ethyl acetate and finally with methanol.
6. Eluent collected was been evaporated using rotary evaporator and the eluent was weighted

The extracted was further subjected to GC-FID analysis

Extraction of Phytochemicals

0.1g of Clove (*Syzgium aromaticum*) extract was weighed and transferred in a test tube and 15ml ethanol and 10ml of 50% m/v potassium hydroxide was added. The test tube was allowed to react in a water bath at 60°C for 3 hrs mins. After the reaction time, the reaction product contained in the test tube was transferred to a separatory funnel. The tube was washed successfully with 20ml of ethanol, 10ml of cold water, 10ml of hot water and 3ml of hexane, which was all transferred to the funnel. The Clove (*Syzgium aromaticum*) extract was combined and washed three times with 10ml of 10% v/v ethanol aqueous solution. The ethanol solvent was evaporated. The sample was solubilized in 1000ul of pyridine of which 200ul was transferred to a vial for analysis.

Quantification by GC - FID

The analysis of phytochemical was performed on a BUCK M910 Gas chromatography equipped with a flame ionization detector. A RESTEK 15meter MXT-1 column (15m x 250um x 0.15um) was used. The injector temperature was 280°C with splitless injection of 2ul of sample and a linear velocity of 30cms⁻¹, Helium 5.0pas was the carrier gas with a flow rate of 40 mlmin⁻¹. The oven operated initially at 200°C, it was heated to 330°C at a rate of 3°C min⁻¹ and was kept at this temperature for 5min. the detector operated at a temperature of 320°C. Phytochemicals were determined by the ratio between the area and mass of internal standard and the area of the

identified phytochemicals. The concentration of the different phytochemicals expresses in ug/g.

Assessment of in Vitro Anti-Oxidant Activity

ABTS Scavenging Effects

Method

The antioxidant effect of the clove was studied using ABTS (2, 2'-azino-bis-3-ethyl benzthiazoline-6-sulphonic acid) radical cation decolorization assay according to the method of Shirwaikar *et al.* (2006).

Principle

Anti-oxidant effect of the clove extract was studied using (2, 2'-azino-bis- 3-ethyl benzthiazoline-6-sulphonic acid) radical cation decolorization assay. The ABTs assay measures the relative ability of anti-oxidant to scavenge the ABTS radical cations (ABTS+) which has a dark blue color is reduced by an antioxidant to give a colorless ABTs product having absorption maxima at 745nm.

Procedure

ABTS radical cations (ABTS+) techniques were produced by reacting ABTS solution (7mM) with 2.45mM ammonium persulphate. The mixture was allowed to stand in the dark at room temperature for 12-16 hours before use. Aliquots (100mg, 200mg and 300mg) of the different samples extract were added to 0.3ml of ABTs solution and final volume was made up to 1000mg with ethanol. The absorbance was read at 745nm in a spectrophotometer (Genesys 10-S, USA) and the per cent scavenging activity was calculated using the formula:

$$\text{Scavenging activity (\%)} = \frac{(\text{Control} - \text{test})}{\text{Control}} \times 100$$

Hydrogen Peroxide Scavenging Effects

Method

The ability of the clove (*Syzgium aromaticum*) to scavenge hydrogen peroxide was assessed by the method of Ruch *et al.* (1989).

Principle

The ability of clove to scavenge hydrogen peroxide. It was determined by measuring decrement of H_2O_2 in an incubation system containing H_2O_2 and the scavenging activity using classical UV-method at 230nm.

Procedure

A solution of H_2O_2 (40mM) was prepared in phosphate buffer. Clove (*Syzygium aromaticum*) extracts at the concentration of (100mg, 200mg and 300mg) were added to a test-tube and the volume was made up to 1000mg respectively. And, H_2O_2 solution (0.6ml) added and the total volume was made up to 3ml by adding 2.3ml of distilled water. The absorbance of the reaction mixture was recorded at 230nm in a spectrophotometer (Genesys 10-S, USA). A blank solution containing phosphate buffer, without H_2O_2 was prepared. The extent of H_2O_2 scavenging of the sample samples was calculated as:

$$\% \text{ scavenging of hydrogen peroxide} = \frac{(A_0 - A_1) \times 100}{A_0}$$

A₀ - Absorbance of control

A₁ - Absorbance in the presence of sample

Assay of Lipid Peroxidation

Method

The extent of lipid peroxidation was estimated according to the method of Okhawa *et al.* (1979).

Principle

Malondialdehyde (MDA) formed from the breakdown of polyunsaturated fatty acids serves as a convenient index for the determination of the extent of peroxidation reaction. MDA, a product of lipid peroxidation reacts with TBA (thiobarbituric acid) to give a pink colored product having absorption maxima at 535nm.

Procedure

Different concentration of the extract (100mg, 200mg and 300mg) were prepared in phosphate buffer. A 20% liver homogenate was prepared in phosphate buffer (pH 7.2). 0.5ml of the homogenate of the extracts and

0.12M phosphate buffer to make up 1000 mg), 1.0ml of TCA and 1.0ml of TBA were added and mixed thoroughly. The mixture was heated in a boiling water bath for 20 minutes. The tubes were centrifuged at 1000g for 20 minutes and the absorbance was read at 535nm in a spectrophotometer (Genesys 10-S, USA) against a blank containing all the reagents except the homogenate. The MDA equivalents of the samples were calculated using the extinction coefficient $1.56 \times 10^5 \text{ M}^{-1}\text{cm}^{-1}$.

DPPH Spectrophotometric Assay

Method

The scavenging ability of the natural antioxidants of the sample towards the stable free radical DPPH was measured by the method of Mensor *et al.* (2001).

Principle

DPPH free radical method is an antioxidant assay based on electron-transfer that produces solution in ethanol. This free radical, stable at room temperature is reduced in the presence of an anti-oxidant molecule, giving rise to colorless ethanol solution.

Procedure

Clove (*Syzygium aromaticum*) extracts at the concentration of (100mg, 200mg and 300mg) were added to a test-tube and the volume was made up to 1000mg respectively. And 0.5ml of 0.1mM methanolic solution of DPPH and 0.48ml of methanol was added. The mixture was allowed to react at room temperature for 30 minutes. Methanol served as the blank and DPPH in methanol, without the clove (*Syzygium aromaticum*) extract, served as the positive control while butylated hydroxytoluene (BHT) served as reference. After 30 minutes of incubation, the discoloration of the purple color was measured at 518nm in a spectrophotometer (Genesys 10-S, USA). The radical scavenging activity was calculated as follows:

$$\text{Scavenging activity \%} = \frac{100 - \frac{A_{518}(\text{sample}) - A_{518}(\text{blank})}{A_{518}(\text{blank})} \times 100}{1}$$

Assessment of *in Vitro* Anti-Inflammatory Activity

Inhibition of Albumin Denaturation

Method

The anti-inflammatory activity of Clove (*Syzygium aromaticum*) was studied by using inhibition of albumin denaturation technique which was studied according to Mizushima et al [1968] and Sakat et al [2010] followed with minor modifications.

Principle

The egg albumin denaturation assay is based on the idea that substance with anti-inflammatory properties has the ability to stabilize protein structure and prevent denaturation which is frequently linked to inflammation and tissue damage.

Procedure

The reaction mixture was consisting of test extracts and 1% aqueous solution of bovine albumin fraction, pH of the reaction mixture was adjusted using small amount of 1N HCl. The homogenate (0.1ml, 0.2ml, 0.3ml of the extracts and distilled water to make up 1 ml) and 1ml of 1% aqueous solution of bovine albumin fraction was added and incubated at 37 °C for 20 min and then heated to 51 °C for 20 min, after cooling the samples the turbidity was measured at 660nm. (UV Visible Spectrophotometer Model 371, Elico India Ltd). The Percentage inhibition of protein denaturation was calculated as follows:

Percentage inhibition = (Abs Control – Abs Sample) X 100/ Abs control.

Heat Induced Haemolysis

Method

According to the method by Sakat *et al*, 2010.

Principle

It involved stabilization of human red blood membrane by hypo tonicity induced membrane lysis.

Procedure

The reaction mixture (2ml) consisted of test sample of different concentrations (100mg,

200mg and 300mg of the extracts and distilled water to make up 1000mg) and 1 ml of 10% RBCs suspension, instead of test sample only saline was added to the control test tube. Aspirin was used as a standard drug. All the centrifuge tubes containing reaction mixture were incubated in water bath at 56 °C for 30min. At the end of the incubation the tubes were cooled under running tap water. The reaction mixture was centrifuged at 2500 rpm for 5 min and the absorbance of the supernatants was taken at 560 nm.

The Percentage inhibition of Hemolysis was calculated as follows:

Percentage inhibition = (Abs control – Abs sample) X 100/ Abs control

Anti-Proteinase Activity

Method

The test was performed according to the modified method of Oyedepo et al, 2012 and Sakat et al 2010

Principle

It is a method that measured proteinase inhibitors ability to reduce tissue damage by proteinase that are released by dead or dying inflammatory cells.

Procedure

The reaction mixture (2 ml) was containing 0.06 mg trypsin, 1 ml 20 mM Tris HCl buffer (pH 7.4) and 1 ml test sample of different concentrations (100mg, 200mg and 300mg of the extracts and distilled water to make up 1000mg). The mixture was incubated at 37°C for 5 min and then 1 ml of 0.8% (w/v) casein was added. The mixture was incubated for an additional 20 min. 2 ml of 70% perchloric acid was added to arrest the reaction. Cloudy suspension was centrifuged and the absorbance of the supernatant was read at 210 nm against buffer as blank. The experiment was performed in triplicate. The percentage inhibition of proteinase inhibitory activity was calculated.

Percentage inhibition = (Abs control – Abs sample) X 100/ Abs control.

Results

Extraction of Phytochemicals

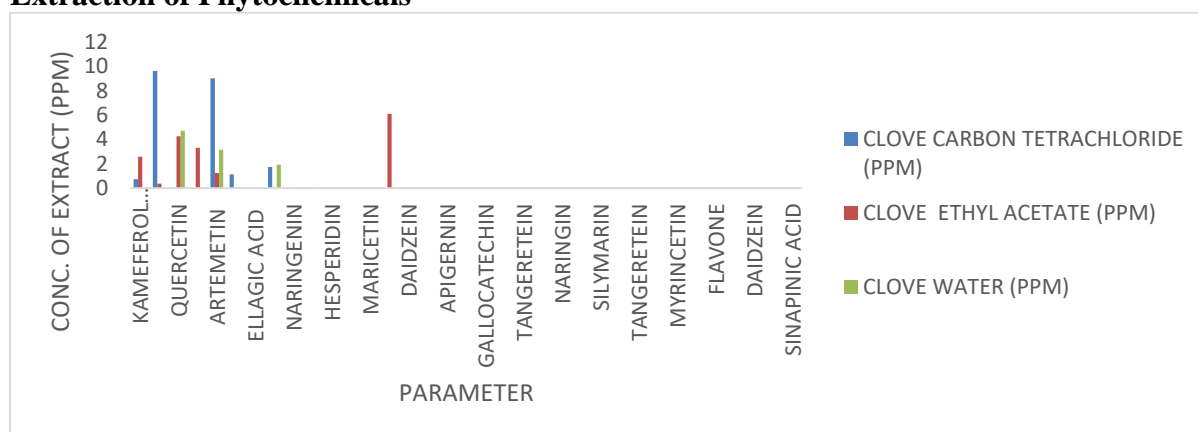


Fig.1: Phytochemical Profile of all the Extracts

This result show that catechin has the highest concentration in carbon tetrachloride, followed by artementin in carbon tetrachloride extracts and epicatechin in ethyl acetate extract with the concentration of 9.61826ppm, 9.01830ppm and 6.08818ppm respectively while naringenin in ethyl acetate, retusin in ethyl acetate and catechin in aqueous extracts has the lowest concentration of phytochemical profile with the concentration of 0.0177608ppm, 0.0181873ppm and 0.0182906ppm respectively when compared

with each other. However, when compared with each other carbon tetrachloride extract has the highest phytochemical composition with the total concentration of 22.311273ppm while aqueous extracts have the lowest phytochemical composition with total concentration of 9.8821797ppm when compared to each other.

Assessment of *in Vitro* Anti-Oxidant Activity

ABTS Scavenging Effect

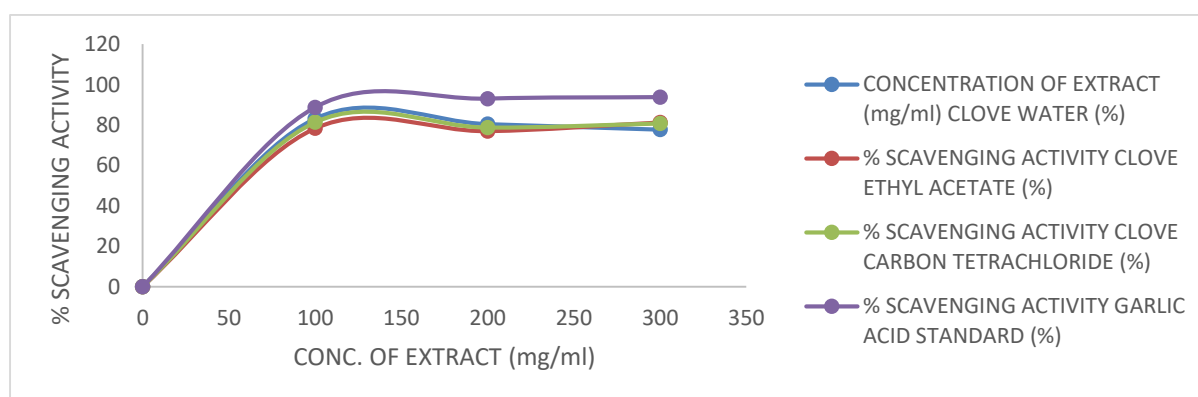


Fig.2: ABTs Percentage Scavenging Effect of all the Extract with Garlic Acid as Standard

The result above shows that there is high carbon tetrachloride of clove ex tract. The percentage scavenging activity of aqueous extract when compared with ethyl acetate and extract is 80.44% while carbon tetrachloride

extract is 80.312% and ethyl acetate extract is 78.86% when compared with standard of 91.83%. As the concentration increases, the result shows that there is increase in the percentage scavenging activity of ethyl acetate

and carbon tetrachloride while there is decrease in the concentration of the aqueous extract. The results also show that all the extracts are significant statistically ($p < 0.05$) when compared with standard.

Mean of ABTs Scavenging Effect

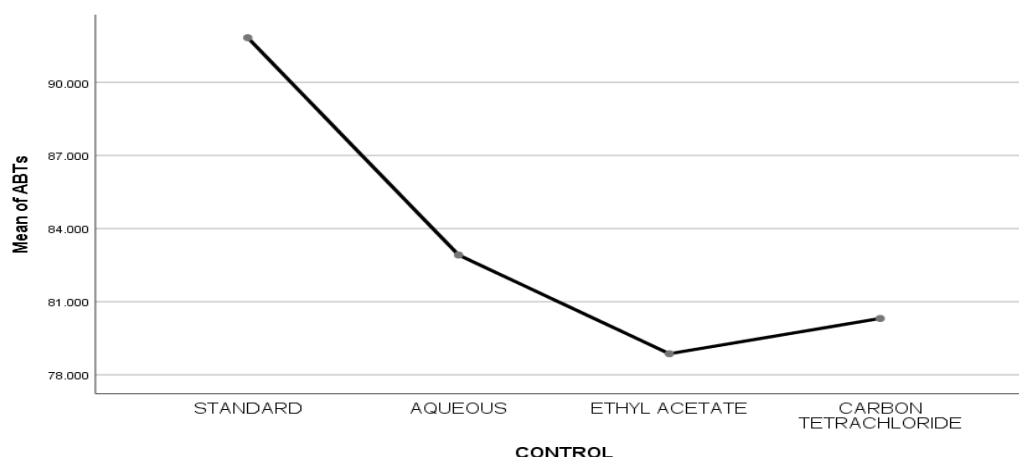


Fig.3: Mean of ABTs Scavenging Effect of Standard, Aqueous, Ethyl Acetate and Carbon Tetrachloride Extracts

The graph shows that all the extracts are significantly low when compared to standard ($p > 0.05$)

Hydrogen Peroxide Scavenging Effects

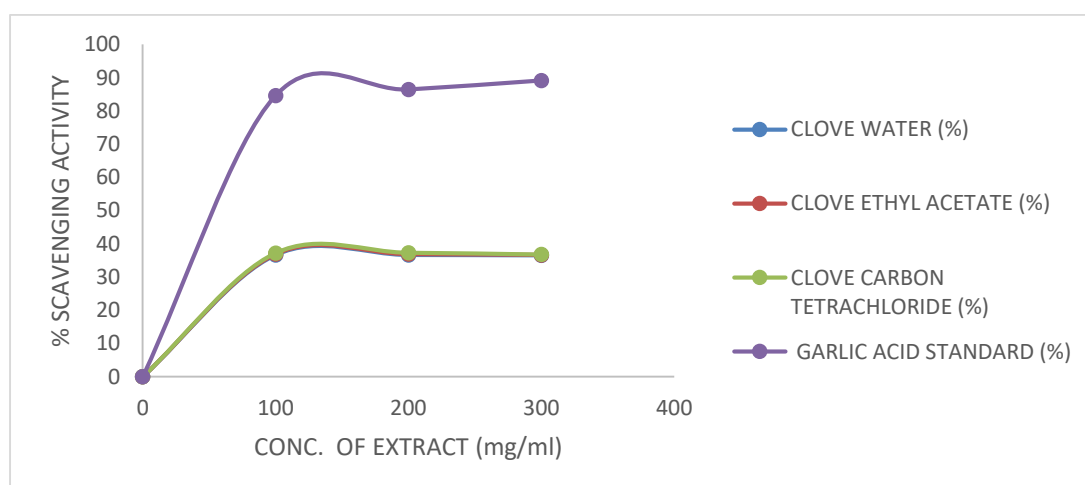


Fig.4: Hydrogen Peroxide Percentage Scavenging Activity of all the Extract with Garlic Acid as Standard.

The fig above shows that carbon tetrachloride extract has the highest percentage scavenging activity when compared to ethyl acetate and clove aqueous extracts. Carbon tetrachloride extracts has the highest average percentage

scavenging activity of 37.096% while ethyl acetate extracts have 36.804% and aqueous extract has 36.623% when compared to standard of 86.732%. As the concentration increase, the result shows that all the extracts

slightly increase in the %scavenging activity having no significant differences.

And it also shows that all the extracts vary significant ($p < 0.05$) when compared with standard.

Means of Hydrogen Peroxide Scavenging Effect

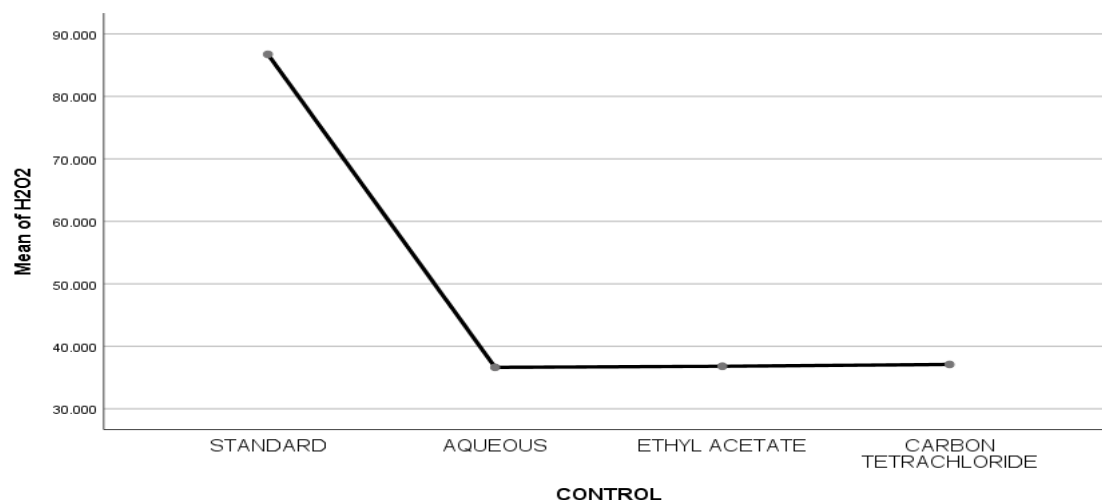


Fig.5: Mean of Hydrogen Peroxide Scavenging Effect of Standard, Aqueous, Ethyl Acetate and Carbon Tetrachloride Extracts

This shows that all the extracts are significantly low when compared to the standard ($p < 0.05$).

Assay of Lipid Peroxidation

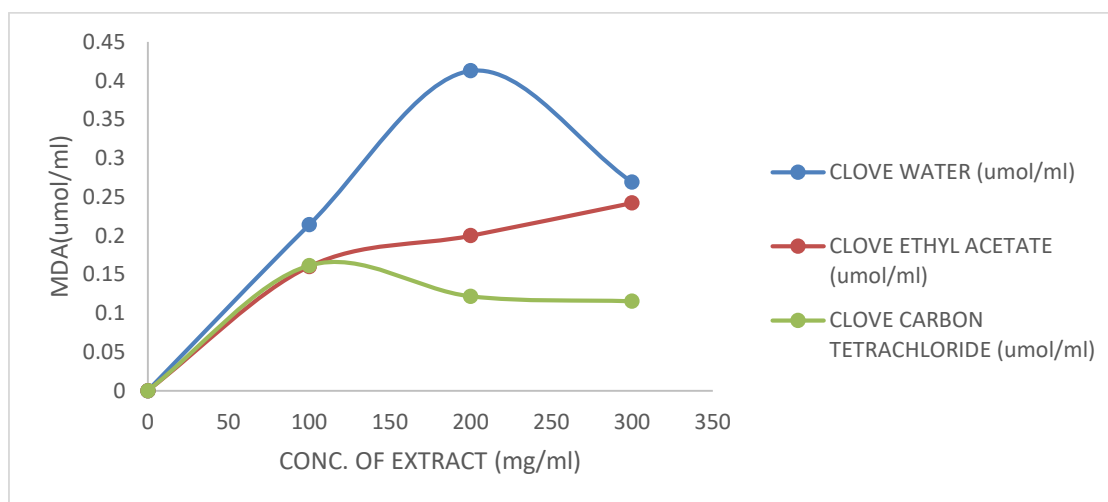


Fig.6: Lipid Peroxidation of All the Extracts

This result shows that aqueous extract has the highest MDA activity when compared to ethyl acetate and carbon tetrachloride extracts. The average MDA activity of aqueous extract is 0.2987 umol/ml while ethyl acetates extract

is 0.2009 umol/ml and carbon tetrachloride extract is 0.1328 umol/ml when compared to each other. There is decrease in MDA activity of carbon tetrachloride extract, while there is increases in MDA activity of ethyl acetate

extract as the concentration of extract decrease of aqueous extract as the increases. Also, there is an increase and concentration of extract exceed 100mg/ml.

DPPH Spectrophotometric Assay

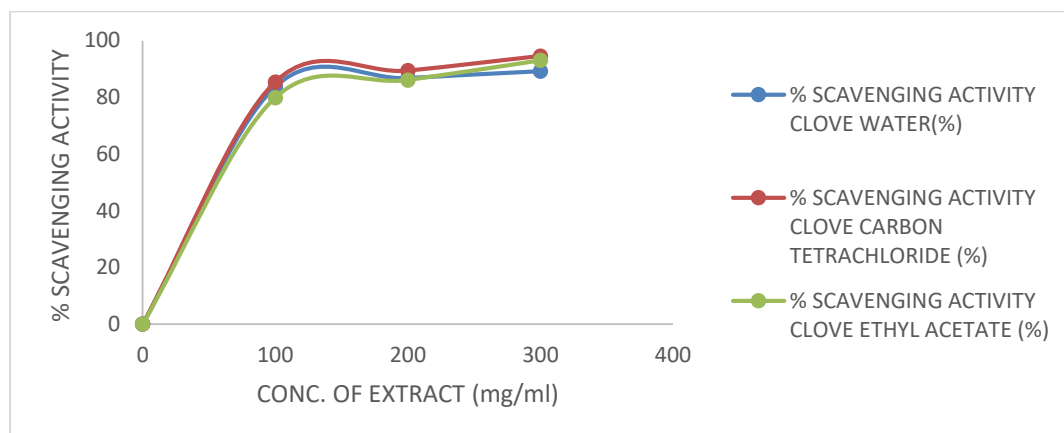


Fig.7: DPPH Spectrophotometric Assay of All the Extract.

The fig above shows that carbon tetrachloride extract has the highest % scavenging activity while ethyl acetates extract has the lowest % scavenging activity when compared to each other. Carbon tetrachloride extract has the average percentage scavenging activity of 89.7966% while aqueous extract of 86.556% and ethyl acetates extract of 86.332% as the

concentration of extract increases. It also shows that there is increase in DPPH % scavenging activity of both ethyl acetate and carbon tetrachloride extracts while aqueous has a decrease in the % scavenging activity as the concentration of the extract increase.

Assessment of *in Vitro* Anti-Inflammatory Activity Inhibition of Albumin Denaturation

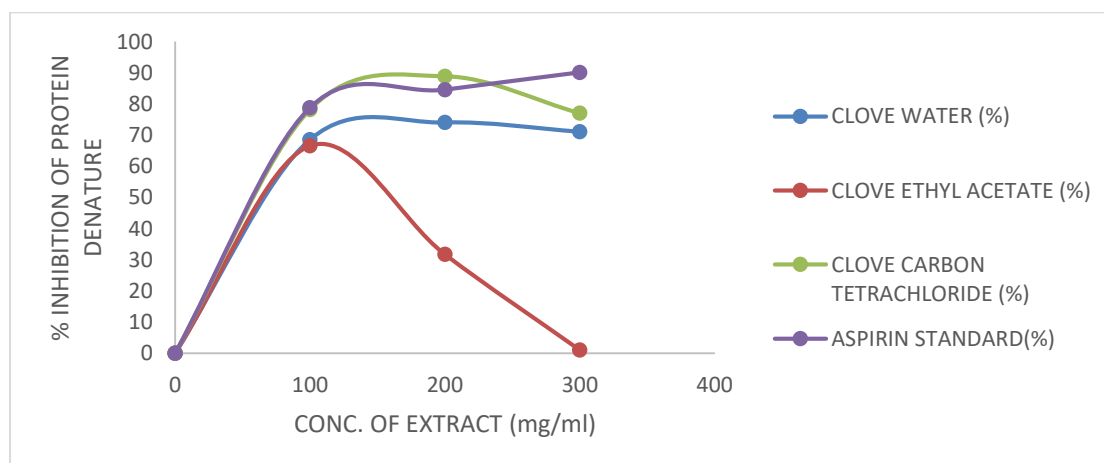


Fig.8: % Inhibition of Albumin Denaturation of all the Extracts with Aspirin as Standard

This graph above shows that carbon albumin denaturation when compared to tetrachloride extract has the highest standard. The average % inhibition of carbon concentration of % inhibition while ethyl tetrachloride extract is 81.3785% while acetate extract has the lowest % inhibition of aqueous extract is 71.33% and ethyl acetate

extract is 33.11% when compared to standard of 84.4727%. It also shows that % inhibition of albumin denaturation of carbon tetrachloride and ethyl acetate decrease as the concentration of extracts exceed 200mg/ml while clove aqueous decreases in % inhibition

as the concentration of extract increases. However, the result also shows that aqueous and carbon tetrachloride extract are not significant statistically ($p < 0.05$) while ethyl acetate extract is significant ($p > 0.05$) when compared to standard.

Mean of %Inhibition of Albumin Denaturation

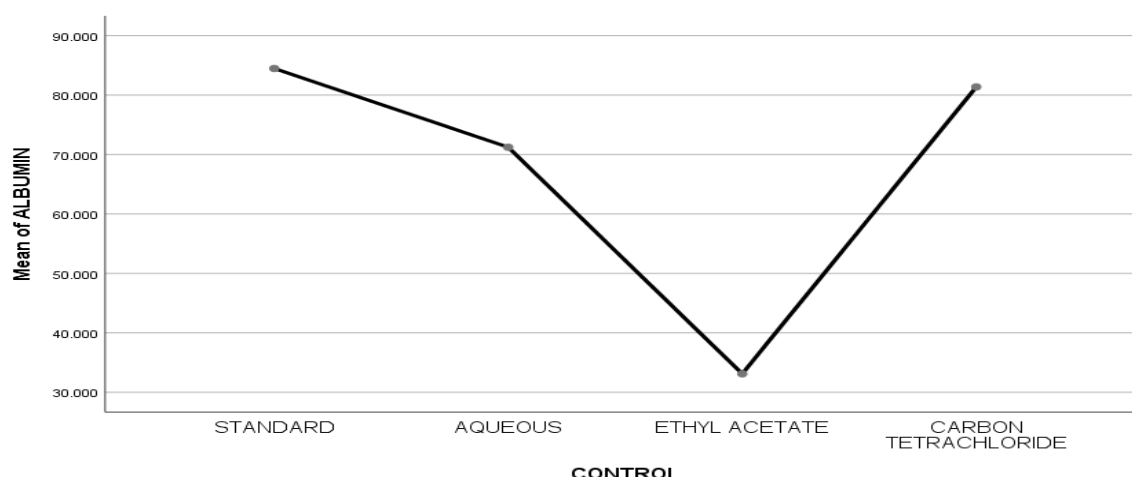


Fig.9: Mean of albumin denaturation inhibition of standard, aqueous, ethyl acetate and carbon tetrachloride extracts

The graph shows that carbon tetrachloride and aqueous extracts are not significantly ($p > 0.05$) while ethyl acetate extract is significantly low ($p < 0.05$) when compared to standard.

Heat Induced Haemolysis

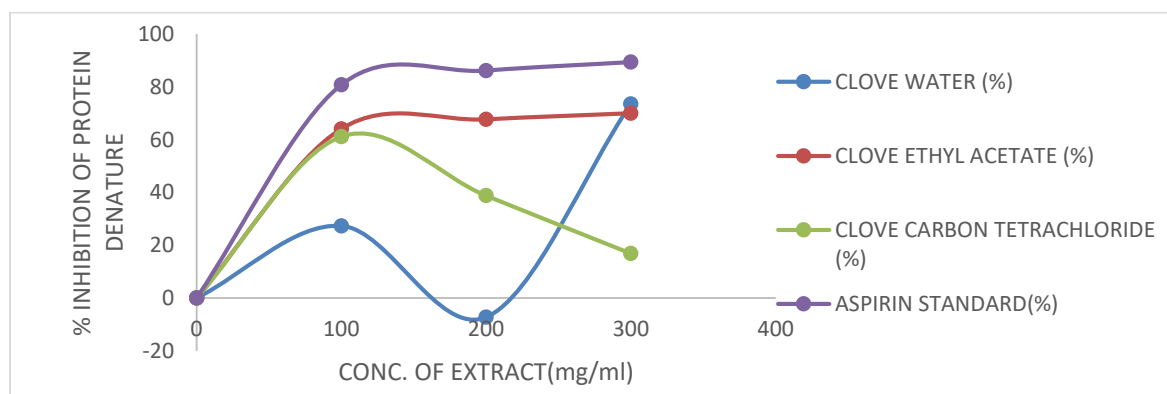


Fig.10: % Inhibition of Heat Induced Haemolysis of All the Extract with Aspirin as Standard

The result show that % inhibition of ethyl acetate extract is high while the % inhibition of aqueous extract is low when compared to standard. It also evaluates that there is an increase in ethyl acetate extract and decrease in % inhibition of carbon tetrachloride extract while aqueous extract show increase and decreases in % inhibition as the concentration of extract increases when compared to standard. The average % inhibition of ethyl

acetate extract is 67.4368% while carbon tetrachloride extract is 38.9419% and aqueous extract is 31.205% when compared to standard of 85.4415%. However, the result also shows

that aqueous and carbon tetrachloride extract are significant statistically ($p < 0.05$) while ethyl acetate extract is not significant ($p > 0.05$) when compared to standard

Mean of Heat Induced Haemolysis

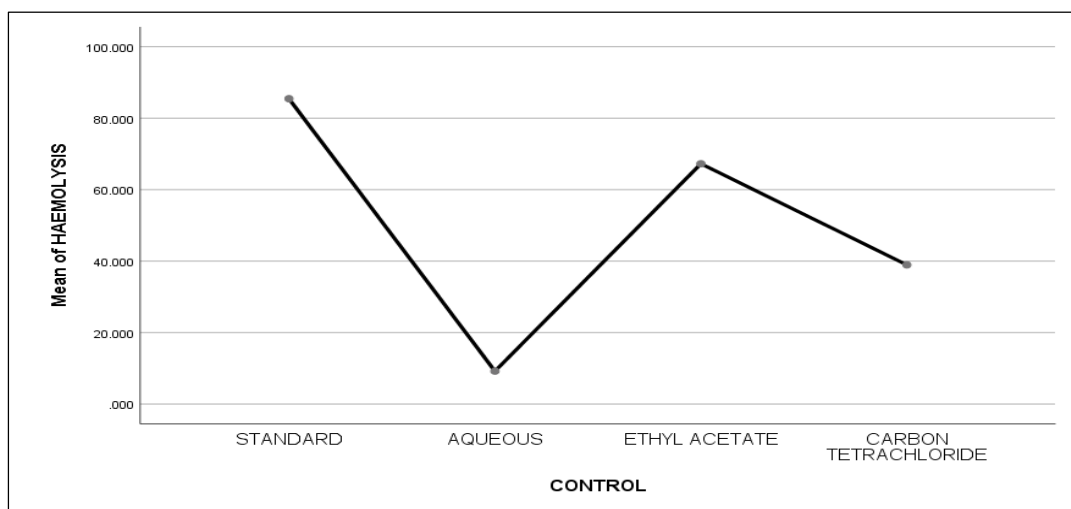


Fig.11: Mean of Heat Induced Haemolysis Inhibition of Standard, Aqueous, Ethyl Acetate and Carbon Tetrachloride

The graph shows that ethyl acetate extract is not significantly ($p > 0.05$) while carbon tetrachloride and aqueous extract are significantly low ($p < 0.05$) when compared to standard.

Anti-Proteinase Activity

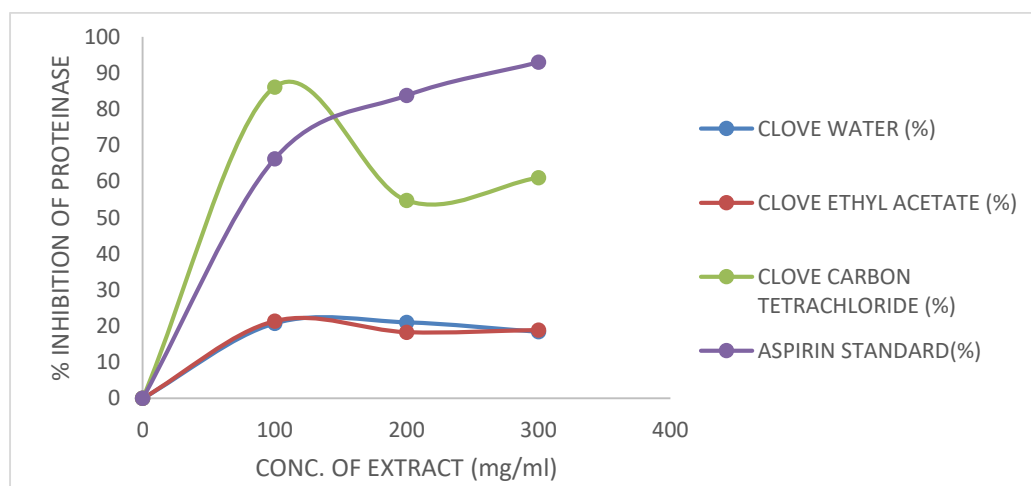


Fig. 12: % Inhibition of Anti-Proteinase Activity of All the Extract with Aspirin as Standard.

The graph above shows that carbon tetrachloride extract has a high % inhibition of 67.3429% and aqueous extract has a low % inhibition of 20.0875% and ethyl acetates extract of 19.5306% average inhibition when compared to standard. As the Carbon tetrachloride extract show an average concentration increases, there is increases

and decreases of % inhibition of all the extract. Moreover, the result also shows that aqueous and ethyl acetate extract are significant ($p < 0.05$) while carbon tetrachloride is not significant ($p > 0.05$) when compared to standard.

Mean of Anti-Proteinase Activity

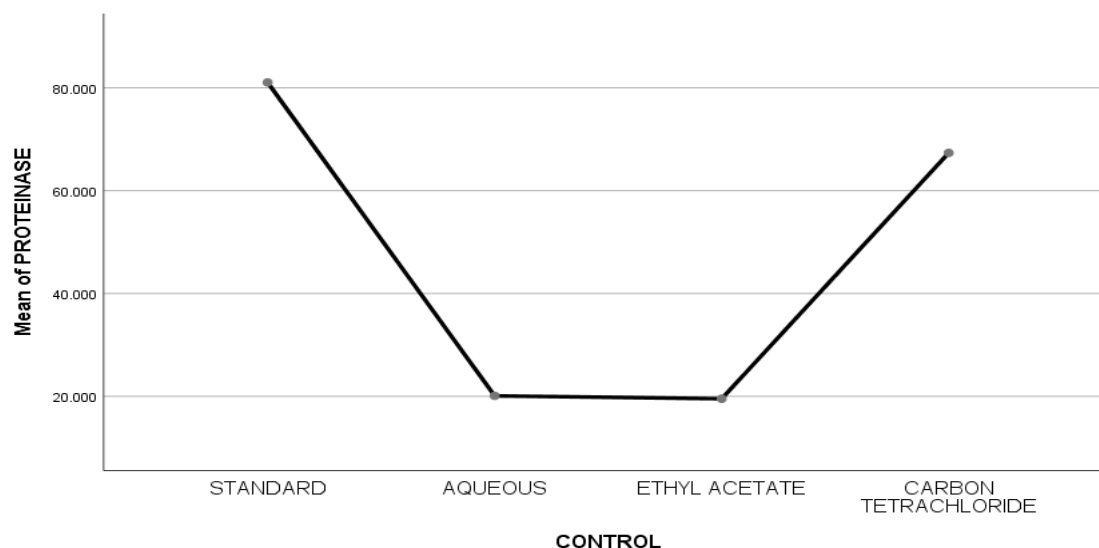


Fig. 13 Mean of Anti-Proteinase Inhibition of Standard, Aqueous, Ethyl Acetate and Carbon Tetrachloride

The graph shows that clove carbon tetrachloride is not significantly ($p > 0.05$) compared to standard. while ethyl acetate and aqueous are significantly low statistically ($p < 0.05$) when compared to standard.

Conclusion

The investigation into the biochemical activities of clove (*Syzygium aromaticum*) extracts reveals both diverse and promising properties. The anti-proteinase activity, lipid peroxidation, hydrogen peroxide scavenging, ABTS scavenging, and DPPH scavenging activities showcase the potential therapeutic applications of clove. The extracts exhibit varied inhibition levels in anti-proteinase activity, distinct antioxidant patterns in lipid peroxidation, and reliable potential in neutralizing hydrogen peroxide radicals. The ABTS and DPPH scavenging activities demonstrate commendable antioxidant potential, emphasizing clove's role as a natural source for the development of antioxidant-rich formulations and therapeutic interventions. The nuanced variations observed underscore the need for further research to elucidate

specific compounds responsible for these activities and optimize extraction methods for enhanced efficacy. Overall, clove emerges as a promising natural resource with multifaceted bioactive potential.

References

- Abdel-Wahhab MA, Aly SE. (2005). Antioxidant property of *Nigella sativa* (black cumin) and *Syzygium aromaticum* (clove) in rats during aflatoxicosis. *J Appl Toxicol.*; 25(3):218–223
- Adams R.P. (2007). Identification of Essential Oil Components by Gas Chromatography/Mass Spectroscopy (4th ed.), Allured Pub. Corp, Carol Stream, Ill

- Adams, M., Efferth T. and Bauer, R. (2006). Activity guided isolation of scopoletin and isoscapoletin, the inhibitory active principles towards CCRF-CEM leukemia cells and multidrug resistant CEM/ADR5000 cells, from *Artemisia argyi*. *Plant Medicine*, **72**: 862-86
- Bamdad F, Kadivar M, Keramat J. (2006). Evaluation of phenolic content and antioxidant activity of Iranian caraway in comparison with clove and BHT using model systems and vegetable oil. *Int J Food Sci Technol.*; 41(Suppl 1): S20–S27
- Adefegha S.A, Oboh G, Adefegha O. M, Boligon A.A and Athayde M. L (2014). Antihyperglycemic, hypolipidemic, hepatoprotective and antioxidative effects of dietary clove (*Syzygium aromaticum*) bud powder in a high-fat diet/streptozotocin-induced diabetes rat model. *Journal of the Science of Food and Agriculture*. <https://doi.org/10.1002/jsfa.6617>
- Arulselvan, P., Fard, M. T., Tan, W. S., Gothai, S., Fakurazi, S., Norhaizan, M. E. and Kumar, S. S. (2016). Role of Antioxidants and Natural Products in Inflammation. *Oxidative Medicine and Cellular Longevity*. 1–15
- Arzel, E., Rocca, P., Grellier, P., Labaeid, M., Frappier, F., Gueritte, F., Gaspard, C., Marsais, F., Godard, A. and Queguiner, G. (2001). New synthesis of benzo-delta-carbolines, cryptolepines, and their salts: *in vitro* cytotoxic, antiplasmodial, and antitrypanosomal activities of delta-carbolines, benzo-delta-carbolines, and cryptolepines. *Journal of Medicinal Chemistry*, 44: 949-960.
- Bacanli M., Dilsiz S. A., Başaran N. and Başaran A. A. (2019). Effects of phytochemicals against diabetes. *Advances in Food and Nutrition Research*. 209–238.
- Batiha G. E. S, Alkazmi L. M, Wasef L. G., Beshbishy A. M, Nadwa E. H, and Rashwan E. K. (2020). “*Syzygium aromaticum* L. (Myrtaceae): traditional uses, bioactive chemical constituents, pharmacological and toxicological activities,” *Biomolecules*, vol. 10, pp. 1–16.
- Batiha G.E.S, Alkazmi LM, Wasef L.G, Beshbishy A.M, Mbaveng A. T and Kaete V (2017). Therapeutic Potential against Metabolic Inflammatory Infectious and Systemic Diseases; 29.
- Caceres, A., Giron, L. M. and Martinez, A. M. (2007). Diuretic activity of plants used for the treatment of urinary ailments in Guatemala. *Journal of Ethnopharmacology*, 19: 233-24
- Chatterjee D, Bhattacharjee P. (2013). Comparative evaluation of the antioxidant efficacy of frying encapsulated and un-encapsulated eugenol-rich clove extracts in soybean oil: shelf-life and stability of soybean oil. *J Food Eng.* 117(4):545–550
- Coe, F. G. and Anderson, G. J. (2006). Ethnobotany in the Garifuna of Eastern Nicaragua. *Economic Botany*, 50: 71-107.

- Cortés-Rojas D.F, de Souza C.R.F, Oliveira W.P. (2014). Clove (*Syzygium aromaticum*): A precious spice. *Asian Pac J Trop Biomed*; 4(2): 90–6.
- Davis C, Henry L, Bernard C, Anders D, Robinson M, Charlotte J, and Grethe H. (2019). Effect of clove (*Syzygium aromaticum*) and seaweed (*Kappaphycus alvarezii*) water extracts pretreatment on lipid oxidation in sun-dried sardines (*Rastrineobola argentea*) from Lake Victoria, Tanzania. *FoodSci Nutr*. 7(4): 1406–1416.
- De Cássia da Silveira e Sá. R, Andrade L. N, and de Sousa D. P, (2013). “A review on anti- inflammatory activity of monoterpenes,” *Molecules*, vol. 18, no. 1, pp. 1227–1254,
- Diego F C, Claudia Regina Fernandes de Souza, and Wanderley Pereira Oliveira (2014). Clove (*Syzygium aromaticum*): a precious spice. *Asian Pac J Trop Biomed*. Feb; 4(2): 90–96.
- Dorsaf B.H., S alma K.E., Rami R., Nessrine G., Rouguiata K., Manef A. and Jalloul B (2021). Clove Buds Essential Oil: The Impact of Grinding on the Chemical Composition and Its Biological Activities Involved Consumer’s Health Security. *Bio Med Research International* Volume 2021, Article ID 9940591, 11 pages
- Dudonné S, Vitrac X, Coutière P, Woillez M and Mérillon J.M (2009). Comparative study of antioxidant properties and total phenolic content of 30 plant extracts of industrial 1774interest using DPPH, ABTS, FRAP, SOD, and ORAC assays. *J Agric Food Chem*; 57(5):1768
- Egigu, M. C, Ibrahim, M. A, Yahya, A and Holopainen, J. K (2010). Yeheb (*Cordeauxia edulis*) extract deters feeding and oviposition of *Plutella xylostella* and attracts its natural enemy. *BioControl*, 55 (5): 613-624.
- Erdemoglu N, Ozkan S and Tosun F. (2007). Alkaloid profile and antimicrobial activity of lupines angustifolius L. Alkaloid extract. *Phytochemistry Reviews*. 6(1):197-201.<https://doi.org/10.1007/s11101-006-9055-8>.
- Gülçin İ. Antioxidant activity of eugenol: a structure-activity relationship study. *J Med Food*. 2011; 14(9):975–985
- Gülçin I, Elmastaş M, Aboul-Enein HY. Antioxidant activity of clove oil-A powerful antioxidant source. *Arab J Chem*. 2012; 5(4):489–499.
- Gülçina İ, Şatb İG, Beydemira Ş, Elmastaş M, Küfrevioğlu Öİ. Comparison of antioxidant activity of clove (*Eugenia caryophyllata* Thunb) buds and lavender (*Lavandula stoechas* L.) *FoodChem*. 2004; 8(3):393–400.
- Halder S, Mehta AK, Kar R, Mustafa M, Mediratta PK, Sharma KK. (2011). Clove oil reverses learning and memory deficits in scopolamine-treated mice. *Planta Med*, 77(8):830–834
- Hazra B., Biswas S., Mandal N. (2008). Antioxidant and free radical scavenging activity of *Spondias pinnata*. *BMC Complementary and Alternative Medicine*, 8:1–10

- Kola.O. A (2019). Phytochemical and antioxidants properties of methanolic extracts opulp, seed, leaf and stem bark of velvet tamarind (*Dialium guineense*) plant. *Journal of Underutilized Legumes.1 (1):159-168*
- Laseve (1996). Mass Spectra and Retention Indice Data Base Université de Québec à Chicoutoumi (UQAC), Canada
- Leelaprakash, G. and Dass, S.M. (2011). *In vitro* Anti-Inflammatory activity of Methanol extract of *Enicostemma Axillare*. *International Journal of Drug Development and Research*, **3**(3):189-196.
- Małgorzata M.P and Kinga S.S. (2021). Comprehensive study on the antioxidant capacity and phenolic **profiles** of black seed and other spices and herbs: **effect** of solvent and time of extraction. *Journal of Food Measurement and Characterization*. 15:4561–4574.
- Medzhitov R. (2010). Inflammation: new adventures of an old flame. *Cell*. 2010; 140:771–776.
- Mehta KD, Garg GR, Mehta AK, Arora T, Sharma AK, Khanna N, et al. (2010). Reversal of propoxur-induced impairment of memory and oxidative stress by 4'-chlorodiazepam in rats. *Naunyn Schmiedeberg's Arch Pharmacol*, 381(1):1–10.
- Nadia Mendoza and Eleazar M. Escamilla Silva (2018). Introduction to Phytochemicals; Secondary Metabolites of plants with Active Principles for Pharmacological Importance. *Intech Open*
- Nizamutdinova T, Dusio G. F and Gasheva O. Y, (2016). “Mast cells and histamine are triggering the NF- κ B-mediated reactions of adult and aged perilymphatic mesenteric tissues to acute inflammation,” *Aging*, vol. 8, no. 11, article 3065, 3090.
- Nwiloh, B.U., A.N., Ibiam, U.A. and Aja, P.M. (2012). Phytochemical composition of *Bryophyllum pinnatum* leaves, *International journal of Advanced Biological research*, 2(4):614-616
- Pathirana H.N, Wimalasena S.H, DeSilva B.C, Hossain S and Gang-Joon H (2019). Antibacterial activity of clove essential oil and eugenol against fish pathogenic bacteria isolated from cultured olive flounder (*Paralichthys olivaceus*). *Slov Vet Res*; 56(1): 31-8.
- Pérez-Jiménez J, Neveu V, Vos F and Scalbert (2010). A. Identification of the 100 richest dietary sources of polyphenols: an application of the phenol-explorer database. *Eur J Clin Nutr.*; 64 (Suppl 3): S112–S120.
- Perry G, Raina A.K, Nonomura A, Wataya T, Sayre L.M and Smith M.A. (2000). How important is oxidative damage? Lessons from Alzheimer's disease. *Free Radical Biol. Med.*, 28; pp. 831-834
- Reslee E.V, Ragavan D. Saranya S, Gayathri D and Gomathi K. (2017). Anti-inflammatory activity of *Syzygium aromaticum* silver nanoparticles: In vitro and in silico study. *Asian Journal of Pharmaceutical and Clinical Research* 10(11):370

- Roe K. (2021). An inflammation classification system using cytokine parameters. *Scandinavian journal of immunology*. **93**(2):1-13.
- Soares J.R, Dinis T.C, Cunha A.P, Almeida L. M (1997). Antioxidant activities of some extracts of *Thymus zygis*. *Free Radic. Res.*, 26 (5); p. 469-478
- Sarrami S, Mohajeri F.A, Sadeghizadeh - Yazdi J, Jambarsang S and Sadrabad E. K. (2023). Chemical Composition and Antioxidant Activity of Clove Essential Oil and its Effect on Stability of Sesame Oil under Accelerated Condition. *Journal of Nutrition and Food Security*. 8 (3): 343 -352
- Shibiru T, Sasikumar J. M. and Egigu M.C. (2022). Effect of Extraction Solvents on Total Polyphenolic Content and Antioxidant Capacity of *Syzygium Aromaticum* L. Flower Bud from Ethiopia. *Biomed Res Int*. 2022: 4568944.
- Shahid, H., Rafia, R., Ayesha, M. and Asma, E.B. (2017). Clove: A review of a precious species with multiple uses. *International Journal of Chemical and Biochemical Science*. 11:129-133
- Shan B, Cai YZ, Sun M, Corke H. (2005). Antioxidant capacity of 26 spice extracts and characterization of their phenolic constituents. *J Agric Food Chem*, 53(20):7749–7759
- Shehwaz Anwar, Abdulmajeed G Almutary, Faris Alrumaihi, Arshad Husain Rahmani, Mohammed A Alsahli, Saleh A Almatroodi, Ahmad Almatroudi and Amjad Ali. Khan (2021). 6-Gingerol, a Major Ingredient of Ginger Attenuates *Diethylnitrosamine*-Induced Liver Injury in Rats through the Modulation of Oxidative Stress and Anti-Inflammatory Activity: 6661937. Doi: 10.1155/2021/6661937
- Silveira e Sá R. Cássia da, Andrade L. N., and de Sousa D. P (2008). “Anti-inflammation activities of essential oil and its constituents from indigenous cinnamon (*Cinnamomum osmophloeum*) twigs,” *Bioresource Technology*, vol. 99, no. 9, pp 3908–3913.
- Stone M. J (2017). “Regulation of chemokine–receptor interactions and functions,” *International Journal of Molecular Sciences*, vol. 18, no. 11, article 2415
- Valko, M., Rhodes, C.J., Moncol, J. and Izakovic, M (2006). Free radicals, metals and antioxidants in oxidative stress-induced cancer. *Biological Interactions Journal*, 160:1-40
- Wright H.L, Moots R.J, Bucknall R.C and Edwards S.W. (2010). Neutrophil function in inflammation and inflammatory diseases *Rheumatology*, 49, pp. 1618-1631, 10.1093/rheumatology/keq045
- Young, S and Woodside, J.V. (2001). Antioxidant in health and disease. *Journal of clinical pathology*, 54:176-86.
- Yunusa S, Yusuf UM, Haruna I (2018) Comparison of Essential Oil of Clove Buds Extracted Using Soxhlet and

- Ultrasonic-Assisted Extraction Zahoor Ahmad Lone and, Navin Kumar Jain
Method (Short Communication),7: 1 (2022). Phytochemical Analysis of
Clove (*Syzygium aromaticum*) Dried
Flower Buds Extract and its
Therapeutic Importance. *Journal of
Drug Delivery and Therapeutics* .12(4-
5):87-92
- Yusuf C. S., Tizhe T. D., James U., Zakawa N.
N. and Wazamda P. J. (2021). Isolation
and control of fungal pathogens
associated with spoiled carica papaya
(l.) Fruit in mubi main market using
aqueous leaf extract of sida acuta. (l.).
*European Journal of Biology and
Biotechnology*. **2**(4):44-46